Structural Features and Biological Activities of Bioactive Compounds from *Fortunella margarita* (Lour.) Swingle: A Review

ZENG Hong-Liang(曾红亮)^(1,2,3);CHEN Pei-Lin(陈培琳)⁽¹⁾; HUANG Can-Can(黄灿灿)^(1,3);SHEN Jin-Ye(沈瑾烨)⁽⁴⁾;CHANG Qing(常青)⁽¹⁾;ZHENG Bao-Dong(郑宝东)
^(1,2,3);ZHANG Yi(张怡)^(1,2,3)

(1) College of Food Science, Fujian Agriculture and Forestry University, Fuzhou 350002, China; (2) Fujian Provincial Key Laboratory of Quality Science and Processing Technology in Special Starch, Fujian Agriculture and Forestry University, Fuzhou 350002, China; (3) China-Ireland International Cooperation Centre for Food Material Science and Structure Design, Fujian Agriculture and Forestry University, Fuzhou 350002, China; (4) College of Life Sciences, Fujian Agriculture and Forestry University, Fuzhou 350002, China

ABSTRACT Fortunella margarita (Lour.) Swingle, commonly known as kumquat, is the smallest citrus fruit. It thrives in southeastern China and is widely cultivated and consumed in the world due to its multiple health benefits. It has been used as an important herbal medicine in traditional Chinese medicine and also as one of the most popular fruits. There are various kinds of bioactive compounds in *F. margarita*, such as polysaccharides, limonoids, essential oils, flavonoids, phenolic acids, vitamins, dietary fiber, etc. In addition, many studies have reported that these bioactive compounds can be used as antioxidant, antimicrobial, hypolipidemic, drosophila lure components in functional foods, pharmaceuticals and daily chemical products due to their biological activities. This review focuses on the structural features and biological activities of polysaccharides, limonoids, essential oils and flavonoids and other bioactive substances from *F. margarita* and their potential applications in food, daily chemical and pharmaceutical industries.

Keywords: Fortunella margarita (Lour.) Swingle; bioactive compounds; structural features; biological activities; application; DOI: 10.14102/j.cnki.0254-5861.2011-1788

1 INTRODUCTION

Fortunella margarita (Lour.) Swingle, known as kumquat or cumquat, originates in the southeastern China and is grown for its delicious fruit in many parts of the world, including Europe, Japan, USA, Puerto Rico, Guatemala, Suriname, Colombia, Brazil,

Australia, South Africa and India^[1]. It is the smallest citrus fruit and is distinguished by the fact that it can be eaten completely, including the peel^[2]. *F. margarita* is the characteristic fruit resource in China and is widely cultivated in Fujian, Zhejiang, Jiangxi, Hunan and Guangxi provinces^[3]. In 2015, its production is over 500,000 tons in China and its output value is up to 10 billion^[4]. It is highly nutritious and contains a lot of bioactive compounds, such as polysaccharides, limonoids, essential oils, flavonoids, phenolic acids, vitamins, dietary fiber, amino acids, etc.^[5, 6]. *F. margarita* has been used to prevent the rupture of blood vessels, reduce the fragility and permeability of blood capillaries and slow the hardening of arteries^[7]. Furthermore, it is used in traditional herbal medicines, especially for the treatment of coughs and colds^[8].

The biological activities of F. margarita are closely related to its sufficient bioactive compounds. Zeng et al. [9] reported the antimicrobial activities of F. margarita were concerned with its polysaccharides. Its polysaccharide fractions also displayed antioxidant activities, pancreatic lipase inhibitory effect and bile acid-binding activities^[10]. Different polysaccharide fractions had various structural features, which resulted in the differences of biological activities. Limonin and nomilin were the main components of the limonoids from F. margarita and their antioxidant activities were investigated by Meng^[11]. Essential oils from *F. margarita* peel and seed were determined by the previous literatures^[12, 13]. These oils displayed different chemical components and biological activities. Zheng et al.[14] and Li et al.[15] investigated the chemical components and the functional properties of flavonoids from F. margarita. In addition, the structural characteristics and functional activities of other bioactive compounds from F. margarita were also studied by some literatures, such as phenolic acids, vitamins, dietary fiber, amino acids and minerals^[16]. All of these bioactivities had something to do with the chemical components and molecular structural features of the bioactive compounds. These basic scientific studies make it possible that the bioactive compounds from F. margarita are utilized as active ingredients in food, pharmaceutical and daily chemical industries.

The objective of the review was to generalize and summarize the information of bioactive compounds from *F. margarita*. To generate summary tables and figures, the bioactivities and chemical structures of bioactive compounds were provided by the original papers. Moreover, the potential applications of bioactive compounds as ingredients in food, daily chemical and

pharmaceutical industries were discussed.

2 CHEMICAL COMPOSITION

E. margarita is highly nutritious, containing a variety of bioactive compounds, such as non-starch polysaccharides, essential oils, limonoids, flavonoids, etc. Table 1 exhibits the chemical composition of *F. margarita*. Some data are provided by USDA Food Composition Databases. It contains 80.85 g of water, 1.88 g of protein, 0.86 g of total lipid, 0.52 g of ash, 15.9 g of carbohydrate, 6.5 g of fiber, and 9.36 g of sugars per 100 g of edible portion^[17]. And it is also rich in minerals, vitamins, carotene, cryptoxanthin, lutein, zeaxanthin, and so on. *F. margarita* displays the multiple health benefits due to its various bioactive compounds. Several studies have reported that the hypocholesteremic and hypolipidemic effect of citrus fruits were attributed to their polysaccharides^[18]. Pectic polysaccharides from citrus fruits inhibited the activity of lipase^[19]. Additionally, the glycemic index was regulated by feeding kumquat juice in mice, which was related to its flavonoid compounds.

3 POLYSACCHARIDES

3.1 Structural properties

Polysaccharides, as the main bioactive compounds from F. margarita, approximately accounted for 12% of dried F. margarita^[4]. The yield of polysaccharides from F. margarita (FMPS) was up to 9.15 \pm 0.13% by ultrasonic-microwave synergistic extraction method^[20], which was increased by 405.52%, 128.18% and 76.64% compared to hot water extraction^[21], ultrasonic-assisted extraction^[22] and microwave-assisted extraction^[9] methods, respectively. FMPS was a macromolecular heteropolysaccharide, containing four kinds of polysaccharide fractions with different concentrations and molecular weights^[10]. Size exclusion chromatography, ultrafiltration, and antisolvent precipitation are the primary methods for the fractionation of macromolecular polymers. The chromatography is a more accurate method for purifying and isolating polysaccharides as compared to ultrafiltration and antisolvent precipitation methods. From Fig. 1, four polysaccharide fractions, named as FMPS1, FMPS2, FMPS3 and FMPS4, were isolated

orderly by DEAE Sepharose CL-6B column and Sephadex G-100 gel column^[10].

Different polysaccharides had variously structural properties. There are also some different structures between crude and purified polysaccharides^[23]. The purified polysaccharide molecules aggregated in the solution by confocal laser scanning microscopy (CLSM) (Fig. 2). The polysaccharide network of purified FMPS was observed and unevenly distributed in the medium and the shape of aggregation was compact and smooth while the molecules of the crude FMPS dispersed in the solution system and there was no network structure. The structural features were also affected by the extraction method. Zeng et al.^[24] investigated the effects of different extraction methods on the molar mass distribution and chain conformation of polysaccharides of *F. margarita*. They found that ultrasonic-assisted and microwave-assisted extraction methods had a significant degradation effect on the molar mass of polysaccharides, while ultrasonic microwave synergistic extraction method had no influence upon the polysaccharides.

Detailed structural features of various polysaccharides are shown in Table 2. All of the polysaccharides had the monosaccharide composition of galactose, galacturonic acid and mannose. FMPS1 and FMPS2 had glucose, and FMPS2 and FMPS3 had arabinose while FMPS1, FMPS3 and FMPS4 had rhamnose^[10]. Moreover, the relative molar percentage of their monosaccharide composition was different. The molecular weights of FMPS, FMPS1, FMPS2, FMPS3 and FMPS4 were $6.192 \times 10^6 \, (\pm 2.59\%)$, $2.572 \times 10^7 \, (\pm 0.517\%)$, $1.755 \times 10^6 \, (\pm 2.009\%)$, $2.563 \times 10^5 \, (\pm 1.784\%)$ and $2.411 \times 10^5 \, (\pm 1.808\%)$, respectively^[24]. FMPS3 and FMPS4 had similar molecular weight, indicating these fractions were not isolated easily by ultrafiltration and antisolvent precipitation^[24]. The glycosidic linkages of FMPS and FMPS3 were mainly β -glycosidic with a small amount of α -glycosidic bonds, while FMPS1 and FMPS2 were mainly α -glycosidic with a small amount of β -glycosidic bonds, as well as FMPS4 was only β -glycosidic linkage. The chain conformation of the polysaccharides in aqueous solution varied. FMPS1 had a tight uniform spherical conformation, FMPS2 had a random coil conformation, whereas FMPS, FMPS3 and FMPS4 displayed highly branched structures.

3. 2 Biological activities

Polysaccharides from many species of plants have important roles in cell-cell communication, cell adhesion and molecular recognition in the immune system^[25]. They have some biological activities, including antioxidant activity, antitumor activity,

hypoglycemic effects, antilipidemic functions, anticancer activity, radioprotection, etc^[4]. The biological activities of polysaccharides from *F. margarita* are shown in Table 2. FMPS displayed antioxidant and antibacterial activities. Zeng et al.^[9] reported FMPS displayed a good antibacterial effect on *Staphyloccocus aureus Rosenbach*. The minimal inhibitory concentrations of polysaccharide against *Staphyloccocus aureus Rosenbach*, *Salmonella*, *Escherichia coli*, *Bacillus subtilis* and *Pseudomonas* were 3.13, 50.00, 12.50, 12.50 and 12.50 mg/mL, respectively. FMPS had a certain capacity on scavenging hydroxyl, superoxide and DPPH radicals, and the antioxidant activities increased with the increasing concentration^[24]. Among these four fractions, FMPS1 and FMPS3 had stronger inhibitory effects on pancreatic lipase, and FMPS1 and FMPS2 had stronger bile acid-binding abilities, as well as FMPS3 and FMPS4 exhibited greater scavenging activities against hydroxyl, superoxide and DPPH radicals^[10].

Moreover, the effects of FMPS on the serum lipid level and antioxidant index of plasma and tissues were investigated in the hyperlipidemia rats^[4]. The result showed the contents of TG, TC, LDL-C and NEFA reduced and HDL-C and LIPA increased significantly by feeding FMPS in the hyperlipidemia rats. Meanwhile, the abilities of SOD, GSH-Px, GST and T-AOC enhanced and the content of MDA reduced by feeding FMPS in the hyperlipidemia rats. There was a certain concentration-response relationship. Moreover, the body weight, liver and spleen index of the hyperlipidemia rats reduced significantly, which was relative to the concentration of FMPS. Histopathological micrographs of hepatic tissue and blood vessel morphology of the hyperlipidemia rats showed the fat deposition in liver cells was reduced and the vascular endothelial cells were protected by feeding FMPS in the hyperlipidemia rats. The result indicated FMPS displayed significant regulatory role on the lipid metabolism disorder of the hyperlipidemia rats. Combination with the hypilipidemic effect *in vitro*, the hypolipidemic mechanism of polysaccharides from *F. margarita* in the hyperlipidemia rats was achieved by increasing the lipase activity, reducing the content of lipid and enhancing the activity of antioxidant enzymes.

3. 3 Structure-bioactivity relationship

The hypolipidemic mechanism, including inhibiting the pancreatic lipase activity, binding bile acid and antioxidant activity, was affected by the preliminary structural characteristics of polysaccharide fractions from *F. margarita* (Fig. 1). The inhibitory effects on pancreatic lipase activity were affected by the monosaccharide composition of the polysaccharide fractions,

especially the pectic polysaccharides^[26]. The ability of polysaccharides to bind bile acid might be related to their anionic, cationic, physical properties, monosaccharide composition and molecular weight^[27]. Glucan could effectively bind bile acids through the molecular interactions with bile salts, and the high viscosity of the polysaccharides had hydrodynamic restrictions on bile acid-binding^[28]. Several factors affected the antioxidant activities of the polysaccharides, including their monosaccharide composition, glycosidic linkage, molecular weight and chain conformation^[29]. FMPS3 were mainly pectic polysaccharides with appropriate molecular weight, β -glycosidic linkage and highly-branched chain conformation in aqueous solution.

4 LIMONOIDS

4. 1 Structural features

Limonoids are a class of highly oxidized triterpenes of secondary metabolites. Limonoids exist in the form of a free ligand and a sugar ligand in citrus fruits, especially fruit peel and seed. At present, more than 300 limonoids have been found, the representatives of which are limonin and nomilin. The acidic limonoids are soluble in water, in which, however, the neutral one is not easy to dissolve^[14]. Limonoids are the main substances that cause bitter taste of citrus fruit juice. The extraction, purification, isolation and structural property of limonoids from *F. margarita* were studied by Meng^[11]. The extracts by supercritical carbon dioxide method were isolated and purified by the recrystallization method. Two crystals were obtained and the molecular structure contained olefin, lactone, vinyl ether, epoxy compounds and groups of -CH₂- and -CH₃ by FT-IT and NMR. The chemical structures of the crystals were characterized by ¹D NMR, ²D NMR and LC-MS/MS^[11]. It was found that the limonoids from *F. margarita* were composed of limonin and nomilin. The fragment analysis of limonin and nomilin from *F. margarita* by secondary mass spectrometry are shown in Tables 3 and 4. These chemical structures were in agreement with the standard substances. And the content of limonin from *F. margarita* was higher than that of nomilin, consistent with the results by Zheng et al.^[14].

4. 2 Biological activities

Limonoids from *F. margarita* are the main active ingredients of anti-cancer. Moreover, they have the activities of anti-inflammatory, anti-anxiety, sedation, regulating cholesterol and

preventing atherosclerosis (Fig. 3)^[30]. They have been used as ingredients in food and pharmaceutical industries due to the health benefits. Li et al.^[31] reported the limonoids from *F. margarita* had the inhibitory effect on DNA oxidation. And the limonoids could reduce the oxidizing reaction of lard and affected the oxidizing rate^[11]. These limonoids had strong antibacterial activities against *Bacillus subtilis*, *Staphylococcus aures*, *Escherichia coli*, *Aspergillus niger*, *Shigella*, *Salmonella*, and *Saccharomyces cervisiae*. The minimum inhibition concentrations were 1.25, 1.25, 1.25, 2.50, 2.50, 2.50 and 5.00 mg/mL, respectively. The acidic condition could promote their antibacterial activities^[11]. Murthy et al.^[32] reported that limonin and limonin glycosides inhibited the proliferation of human colon cancer cells. Patil et al.^[33] studied the anticancer activities of five purified limonoids. Studies had shown that it had a significant inhibition of pancreatic cancer cells, which was consistent with Zhang's research^[34]. It is reported by Hafeeze et al.^[35] that the limonoids had insecticidal effect and resistance capacity to aedes.

5 ESSENTIAL OILS

5. 1 Chemical components

Essential oils are aromatic and volatile liquids extracted from various plant as secondary metabolites. *F. margarita* peel and seed contain a large number of oil cells which contain pigment and aromatic oil. When they get ripe, the content of the essential oil from *F. margarita* peel is generally $22\% \sim 28\%$ [36]. The aromatic oil has great quality, which is an important and popular natural chemical raw material and edible flavor. The peel essential oil can be extracted by the methods of squeezing, hydro-distillation, oil separation, carbon dioxide extraction and continuous subcritical water extraction. The essential oils from *F. margarita* peel were extracted by microwave, ultrasonic and supercritical CO₂ fluid methods, and the highest yield was up to 5.08%. The physicochemical properties of the peel essential oil were as follows: 0.4668 of acid value, 4.2456 of ester value, 0.8380 g/mL of density, 1.4707 of diopter, and 1.8920 of optical rotation [37]. The component analysis of peel essential oil was studied by Wang et al. [38]. There are 88 components in peel essential oil, including alkanes, alkenes, alcohols, acids, ketones, aldehydes and ester substances. Essential oils from *F. margarita* peel were mainly composed of D-limonene, myrcene and β -pinene with relative amounts of 72.90%, 6.88% and 3.60% [13].

Moreover, the physicochemical properties and chemical components of seed oil from F. margarita were investigated by Xie et al.^[13]. The physicochemical properties were as below: 200.00 ± 3.45 of saponification value, 130.12 ± 2.67 of iodine value, 1.47 of refractive index, 0.92 of proportion, 1.58 ± 0.06 of acid value and 5.02 ± 0.14 of peroxide value. The chemical components were also detected by GC-MS. The main components were linoleic acid (47.82%), oleic acid (17.06%) and methyl palmitate acid (15.88%)^[39]. The unsaturated fatty acid was up to 64.88%. Its linoleic acid content was generally higher as compared to the rapeseed oil, linseed oil, peanut oil and sesame oil.

5. 2 Biological activities

The biological activities of essential oils from *F. margarita* peel and seed are shown in Fig. 4. The antimicrobial activities of peel essential oil showed that it had the inhibiting effect against *staphylococcus aureus*, *escherichia coli*, *salmonella*, *aspergillus niger* and *yeast*. The inhibiting effect was promoted by adding some acid solution, but the antimicrobial activities had thermal instability^[12]. The peel essential oil displayed the effectively antioxidant activity in edible fat, especially in olive and lard oils. The peroxide values of olive and lard oils decreased with the addition of peel essential oil^[38]. Interestingly, the peel essential oil had great drosophila lure effect by Wang et al.^[38]. When the test time was 40 min, 10 μ L/mL of peel essential oil displayed the strongest drosophila lure effect and the luring rate was up to 81.0%. It can be used as a potential attractive substance in agriculture. In addition, its seed oil displayed strong scavenging abilities against hydroxyl and superoxide radicals and had the great total antioxidant activity. The scavenging ability against hydroxyl radical was stronger as compared to V_c ^[12].

6 FLAVONOIDS

6.1 Chemical components

Flavonoids are plant secondary metabolites and exist in higher plants. Flavonoids usually exist in the form of aglycon and glycosides (one or more glycosyl groups on the ring). Flavonoids are widely found in the peel and pulp from F. margarita. Zheng^[14] used UPLC-PDA-MS to determine the content of 9 flavonoid compounds, such as apigenin 8-C-rutinoside, hesperidin, neohesperidin, rhoifolin, poncirin, acacetin 7-O-rutinoside, phloretin, phloretin 3',5'-di-C- β -glycopyranoside,

apigenin, acacetin 8-C-neohesperidoside and acacetin 6-C-neohesperidoside. It was found that phloretin 3',5'-di-C- β -glycopyranoside was the main component of flavonoids from *F. margarita*. Acacetin 3,6-di-*C*-glucoside, vicenin-2, lucenin-2,4'-methyl ether, narirutin 4'-*O*-glucoside and apigenin 8-*C*-neohesperidoside were identified for the first time in kumquat juice by Barreca et al. [40]. According to literatures, the flavonoids from *F. margarita* were mainly composed of rhoifolin, phloretin 3',5'-di-C-glucoside, vicenin-2, narirutin 4'-O-glucoside, acacetin 8-C-neohesperidoside, and didymin (Fig. 5)[41]. These main components affected the biological activities of lavonoids from *F. margarita*.

6. 2 Biological activities

Flavonoids have an important health effect on the human body. Studies have shown that flavonoids have good antioxidant activity, cardiovascular disease prevention, anti-cancer activity, anti-inflammatory, antibacterial activity, antiviral activity, free anticoagulant, antithrombotic, etc.^[42]. Li et al.^[43-45] reported that flavonoids from *F. margarita* had significant antioxidant effects and promoted the immune function of mice, as well as significantly relieved symptom of diabetes in mice. Chen et al.^[46] reported that the total flavonoids could protect the liver of acute alcoholism of mice. Antioxidant tests of flavonoids from kumquat showed that the inhibition rates of flavonoids against DPPH · and ABTS · were up to 79% and 93%, respectively^[40].

7 OTHER BIOACTIVE COMPOUNDS

In addition to polysaccharides, limonoids, essential oils and flavonoids, *F. margarita* is also rich in phenolic acids, vitamins, dietary fiber and other bioactive substances. Phenolic acid is a compound in form of the same benzene ring with a number of phenolic hydroxyl groups. It is an important secondary metabolite in the plant, which is the second major metabolite of the plant followed by the flavonoids. Most of the phenolic acids in *F. margarita* are the hydroxylated derivatives of cinnamic and benzoic acids^[47]. The coumaric, chlorogenic, caffeic, erucic and ferulic acids from *F. margarita* in Taiwan were determined using HPLC by Wang et al.^[48]. They found the ferulic and erucic acids were the most important soluble phenolic acids in *F. margarita* peel. Some studies had shown that *F. margarita* phenolic acids had various biological activities, such as anti-virus, anti-inflammatory, anti-allergy, immune function antibacterial activity, pest

control effect, etc. *F. margarita* peels contained the high total amounts of lutein, zeaxanthin, β -cryptoxanthin and β -carotene, which exhibited antioxidant activity, anti-cancer activity, enhancing immune function, preventing osteoporosis, preventing night blindness, preventing cardiovascular disease, anti-aging activity, inhibiting tumor cells growth, and so on^[48]. *F. margarita* also contained a certain amount of dietary fiber, which could increase the activity of metabolic granules in order to prevent constipation^[16].

8 APPLICATIONS

So far, *F. margarita* is mainly used as an edible fresh fruit and is rarely processed for products. Its products only contain fruit juice, concentrated juice, jam, wine, vinegar, dried fruit, etc. The comprehensive development and utilization of bioactive compounds from *F. margarita* have become an important direction for the future development of *F. margarita* industry, especially polysaccharides, limonoids, essential oils and flavonoids. FMPS and FMPS3 can be used as novel natural hypolipidemic and antioxidant agents in food industry, respectively. The limonoids and flavonoids can be utilized as natural antibacterial and antioxidant agents in functional food and pharmaceutical industry, respectively. The essential oils have been developed as raw materials in Chinese daily chemical industry, such as sunscreen, skin cream, hand cream and soap.

9 CONCLUSION

In this review, when going through the literatures, it was observed that there are sufficient amount of bioactive compounds in *F. margarita*. The structural features of bioactive compounds contributed to their biological activities. The polysaccharide was a macromolecular heteropolysaccharide, containing four kinds of polysaccharide fractions with different molecular weight. Different polysaccharide fractions displayed antibacterial, antioxidant activities and hypolipidemic effect. The limonoids were composed of limonin and nomilin, which had anti-cancer activity, anti-inflammatory activity, anti-anxiety activity, sedation effect, cholesterol regulation and prevention atherosclerosis. Essential oils from *F. margarita* peel were mainly

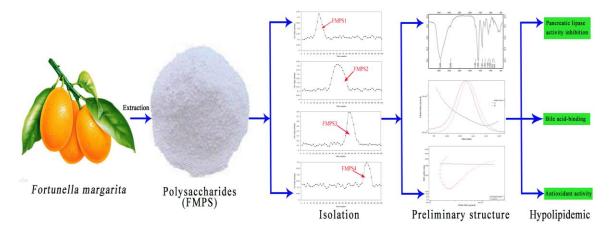
composed of D-limonene, myrcene and β -pinene while linoleic acid, methyl palmitate and oleinic acid were the major components of F. margarita seed oils. Essential oils displayed the antimicrobial, antioxidant activities and drosophila lure effect. The flavonoids were mainly composed of vicenin-2, narirutin 4'-O-glucoside, phloretin 3',5'-di-C-glucoside, rhoifolin, acacetin 8-C-neohesperidoside, didymin, which had a strong capacity to eliminate free radicals. Therefore, evidence suggests that bioactive compounds from F. margarita have potentials as active ingredients for preparing various functional foods, pharmaceutical and daily chemical products due to their valuable biological functions and beneficial health effects.

REFERENCES

- (1) Ag &cs, A.; Nagy, V.; Szab & Z.; M &rk, L.; Ohmacht, R.; Deli, J. Comparative study on the carotenoid composition of the peel and the pulp of different citrus species. *Innov. Food Sci. Emerg.* **2007**, 8, 390-404.
- (2) Peng, L.; Sheu, M.; Lin, L.; Wu, C.; Chiang, H.; Lin, W.; Lee, M.; Chen, H. Effect of heat treatments on the essential oils of kumquat (Fortunella margarita Swingle). Food Chem. 2013, 136, 532-537.
- (3) Zheng, Y. N. Study on the Technologies of Kumquat Juice Concentrating Process. Fujian Agriculture And Forestry University 2012.
- (4) Zeng, H. L. Structural Characterization and Hypolipidemic Mechanism of Polysaccharides from Fortunella Margarita (Lour.) Swingle. Fujian Agriculture And Forestry University 2015.
- (5) Ramful, D.; Tarnus, E.; Aruoma, O. I.; Bourdon, E.; Bahorun, T. Polyphenol composition, vitamin C content and antioxidant capacity of Mauritian citrus fruit pulps. Food Res. Int. 2011, 44, 2088-2099.
- (6) Wu, H. Q. Study on Kumquat Juice Processing Technology with Two Kinds of Enzymatic Method. Fujian Agriculture And Forestry University 2011.
- (7) Abirami, A.; Nagarani, G.; Siddhuraju, P. In vitro antioxidant, anti-diabetic, cholinesterase and tyrosinase inhibitory potential of fresh juice from Citrus hystrix and C. maxima fruits. *Food Sci. Human Well* **2014**, 3, 16-25.
- (8) Zeng, H. L. Extraction, Isolation, Purification, Antibacterial and Antioxidative Activity of Polysaccharides from Fortunella margarita. Fujian Agriculture And Forestry University 2012.
- (9) Zeng, H. L.; Huang, C. C.; Chang, Q.; Xu, W. Y.; Zhang, Y.; Zheng, B. D. Optimization of microwave-assisted extraction of polysaccharide from *Fortunella margarita* (Lour.) Swingle and its antibacterial effect. *Food Machin.* **2016**, 32, 154-160.
- (10) Zeng, H. L.; Miao, S.; Zhang, Y.; Lin, S.; Jian, Y.; Tian, Y. T.; Zheng, B. D. Isolation, preliminary structural characterization and hypolipidemic effect of polysaccharide fractions from *Fortunella margarita* (Lour.) Swingle. *Food Hydrocolloid.* **2016**, 52, 126-136.
- (11) Meng, P. Study on Extraction, Purification, Structural Identification and Biological Activities of Limonoids from Kumquat. Fujian Agriculture And Forestry University 2013.
- (12) Wang, S. Y. Studies on the Extraction and Function of Essential Oil from Kumquat Peel. Fujian Agriculture And Forestry University 2013.
- (13) Xie, J. F. Studies on the Extraction Technology and Components of Fortunella margarita Seed Oil. Fujian Agriculture And Forestry University 2011.
- (14) Zheng, J. Nutritional and Functional Compounds of Major Varieties of Kumquat (Fortunella Swingle) in China. Southwestern University 2015.
- (15) Li, J. L. The Research on Extration and Bioactivities of Active Components in Kumquat. Central South University of Forestry and

- Technology 2007.
- (16) Wang, Y. C.; Chuang, Y. C.; Ku, Y. H. Quantitation of bioactive compounds in citrus fruits cultivated in Taiwan. *Food Chem.* **2007**, 102–1163-1171
- (17) Meng, P. Study actuality of kumquat and its exploitive foreground. Academ. Period. Farm Prod. Proc. 2009, 35-37.
- (18) Chau, C. F.; Huang, Y. L.; Lin, C. Y. Investigation of the cholesterol-lowering action of insoluble fibre derived from the peel of Citrus sinensis L. cv. Liucheng. *Food Chem.* **2004**, 87, 361-366.
- (19) Espinal, R. M.; Parada, A. F.; Restrepo, S. L. P.; Narvázz, C. C. E. Inhibition of digestive enzyme activities by pectic polysaccharides in model solutions. *Bioact. Carboh. Diet. Fibre* **2014**, 4, 27-38.
- (20) Zeng, H. L.; Zhang, Y.; Lin, S.; Jian, Y. Y.; Miao, S.; Zheng, B. D. Ultrasonic-microwave synergistic extraction (UMSE) and molecular weight distribution of polysaccharides from Fortunella margarita (Lour.) Swingle. Sep. Purif. Technol. 2015, 144, 97-106.
- (21) Zeng, H. L.; Lu, X.; Bian, Z. Y.; Lin, Y. F.; Zhang, Y. Optimization of the extraction technique of *Fortunella margarita* polysaccharides via response surface analysis. *J. Fujian Agr. For. Univ. (Nat. Sci. Edit.)* **2012**, 41, 315-319.
- (22) Zeng, H. L.; Zhang, Y.; Zhao, Y. T.; Tian, Y. T.; Miao, S.; Zheng, B. D. Extraction optimization, structure and antioxidant activities of *Fortunella margarita* Swingle polysaccharides. *Int. J. Biol. Macromol.* **2015**, 74, 232-242.
- (23) Zeng, H. L.; Zhang, Y.; Jian, Y. Y.; Tian, Y. T.; Miao, M.; Zheng, B. D. Rheological properties, molecular distribution, and microstructure of *Fortunella margarita* (Lour.) Swingle polysaccharides. *J. Food Sci.* 2015, 80, 742-749.
- (24) Zeng, H. L.; Zhang, Y.; Liu, J.; Zheng, B. D. Molar mass distribution and chain conformation of polysaccharides from *Fortunella margarita* (Lour.) Swingle. *Chin. J. Struct. Chem.* **2014**, 33, 1245-1252.
- (25) Mu, R. J.; Pang, J.; Yuan, Y.; Tan, X. D.; Wang, M. Chen, H.; Chiang, W. T. Progress on the structures and functions of aerogels. Chin. J. Struct. Chem. 2016, 35, 487-497.
- (26) Huang, Y. L.; Chow, C. J.; Tsai, Y. H. Composition, characteristics, and in-vitro physiological effects of the water-soluble polysaccharides from Cassia seed. *Food Chem.* **2012**, 134, 1967-1972.
- (27) Fijan, R.; Basile, M.; Šostar, T. S.; Žagar, E.; Žigon, M.; Lapasin, R. A study of rheological and molecular weight properties of recycled polysaccharides used as thickeners in textile printing. *Carbohyd. Polym.* **2009**, 76, 8-16.
- (28) Gunness, P.; Flanagan, B. M.; Gidley, M. J. Molecular interactions between cereal soluble dietary fibre polymers and a model bile salt deduced from ¹³C NMR titration. *J. Cereal Sci.* **2010**, 52, 444-449.
- (29) Jahanbin, K.; Gohari, A. R.; Moini, S.; Emam-Djomeh, Z.; Masi, P. Isolation, structural characterization and antioxidant activity of a new water-soluble polysaccharide from Acanthophyllum bracteatum roots. *Int. J. Biol. Macromol.* **2011**, 49, 567-572.
- (30) Khalaf, A.; Moore, G. A.; Jones, J. B.; Gmitter Jr, F. G. New insights into the resistance of Nagami kumquat to canker disease. *Physiol. Mol. Plant P* **2007**, 71, 240-250.
- (31) Li, J. L.; Zhang, H.; Zeng, C. Z.; Li, Z. H. Study on ultrasonic extraction technology of limonin from the kumquat. *J. Chin. Instit. Food Sci. Technol.* **2009**, 9, 96-102.
- (32) Murthy, K. N. C.; Jayaprakasha, G. K.; Kumar, V.; Rathore, K. S.; Patil, B. S. Citrus limonin and its glucoside inhibit colon adenocarcinoma cell proliferation through apoptosis. *J. Agr. Food Chem.* **2011**, 59, 2314-2323.
- (33) Patil, J. R.; Jayaprakasha, G. K.; Murthy, K. N. C.; Chetti, M. B.; Patil, B. S. Characterization of Citrus aurantifolia bioactive compounds and their inhibition of human pancreatic cancer cells through apoptosis. *Microchem. J.* **2010**, 94, 108-117.
- (34) Zhang, J. J.; Luo, G.; He, L.; Zhou, L. M. Inhibiting effects of limonin on human hepatocarcinoma cells SMMC-7721 in vitro. Sichuan J. Physiol. Sci. 2007, 29, 157-160.
- (35) Hafeez, F.; Akram, W.; Shaalan, E. A. Mosquito larvicidal activity of citrus limonoids against Aedes albopictus. *Parasitol. Res.* **2011**, 109, 221-229.
- (36) Su, D. L.; Shan, Y. Review of physiologically-active compounds in citrus peels. Mod. Food Sci. Technol. 2006, 22, 260-262.
- (37) Fu, W. Q.; Wang, S. Y.; Zheng, B. D.; Zeng, S. X.; Zhang, Y. Optimization of supercritical CO₂ fluid extraction technology of essential oil from kumquat peel and its physicochemical properties. *Chin. J. Trop. Agr.* **2015**, 35, 55-59.
- (38) Wang, S. Y.; Zhang, Y.; Zheng, B. D. The extraction process of essential oil from Fortunella peel via steam distillation combined

- with microwave and its influence on the attracted activity of the fruit fly. J. Chin. Inst. Food Sci. Technol. 2014, 14, 37-44.
- (39) Zhang, Y.; Xie, J. F.; Zeng, S. X.; Zheng, B. D. Study on the extraction technology and components of *Fortunella Margarita* seed oil by ultrasonic. *J. Chin. Inst. Food Sci. Technol.* **2013**, 13, 35-42.
- (40) Barreca, D.; Bellocco, E.; Caristi, C.; Leuzzi, U.; Gattuso, G. Kumquat (*Fortunella japonica* Swingle) juice: flavonoid distribution and antioxidant properties. *Food Res. Int.* **2011**, 44, 2190-2197.
- (41) Kumamoto, H.; Matsubara, Y.; Iizuka, Y.; Okamoto, K.; Yokoi, K. Structure and hypotensive effect of flavonoid glycosides in orange (Citrus sinensis OSBECK) peelings. *Agr. Biol. Chem.* **1986**, 50, 781-783.
- (42) Lou, S. N.; Lai, Y. C.; Huang, J. D.; Ho, C. T.; Ferng, L. H. A.; Chang, Y. C. Drying effect on flavonoid composition and antioxidant activity of immature kumquat. *Food Chem.* **2015**, 171, 356-363.
- (43) Li, J. L.; Cui, P. W.; Wu, Y. H.; Zeng, C. Z.; Liu, Z. M. Effect of kumquat flavonoids on the anti-oxidation in mice. *Lishizhen Med. Mat. Med. Res.* 2009, 20, 1031-1032.
- (44) Li, J. L.; Zhang, H.; Zeng, C. Z.; Li, Z. H. Study on extraction, purification, structural identification and biological activities of limonoids from kumquat. *J. Chin. Inst. Food Sci. Technol.* **2009**, 9, 96-102.
- (45) Li, J. L.; Li, Z. H.; Zhong, H. Y.; Liu, Z. M. Effect of kumquat flavone on blood glucose in mice. *Pharmacol. Clin. Chin. Mat. Med.* **2007**, 23, 42-44.
- (46) Chen, Z. Y. Study on the anti-Alcoholism and Protecting Liver Effects of Kumquat Component. Hunan Agricultural University 2012.
- (47) Hayat, K.; Zhang, X.; Farooq, U.; Abbas, S.; Xia, S.; Jia, C.; Zhong, F.; Zhang, J. Effect of microwave treatment on phenolic content and antioxidant activity of citrus mandarin pomace. *Food Chem.* **2010**, 123, 423-429.
- (48) Wang, Y. C.; Chuang, Y. C.; Hsu, H. W. The flavonoid, carotenoid and pectin content in peels of citrus cultivated in Taiwan. *Food Chem.* 2008, 106, 277-284.



 ${\bf Fig.\,1.} \quad {\bf Structural\,\, characterization\,\, and\,\, hypolipidemic\,\, effect\,\, of\,\, polysaccharide}$

fractions from F. $margarita^{[10]}$. Reprinted with permission

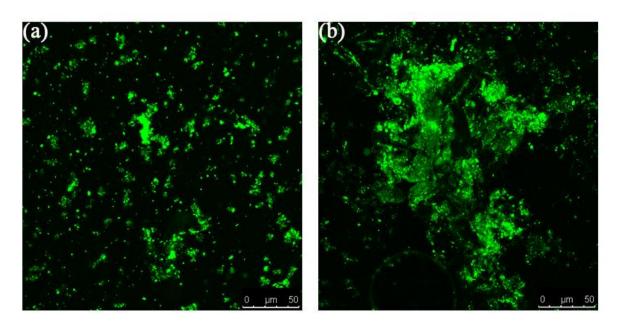


Fig. 2. CLSM images of FMPS solutions: (a) Crude FMPS; (b) Purified FMPS^[23].

Reprinted with permission

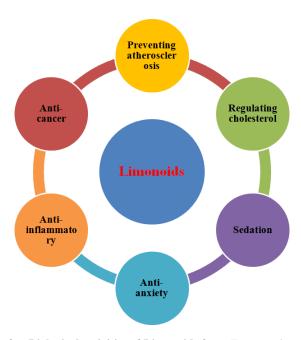


Fig. 3. Biological activities of Limonoids from F. margarita

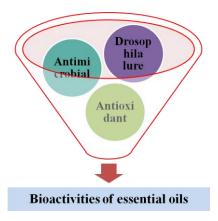


Fig. 4. Biological activities of essential oils from F. margarita

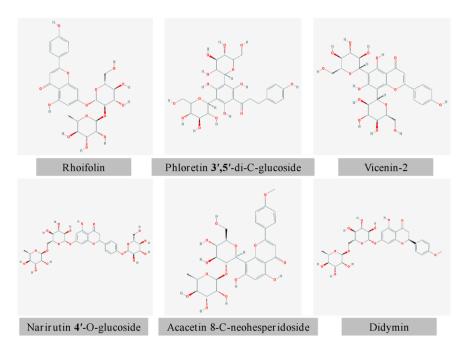


Fig. 5. Chemical components of flavonoids from F. margarita

Table 1. Chemical Composition of F. margarita

| Nutrient | Units | Value per 100 g of edible portion | |
|----------------------|-------|-----------------------------------|--|
| Water | g | 80.85 | |
| Protein | g | 1.88 | |
| Total lipid (fat) | g | 0.86 | |
| Ash | g | 0.52 | |
| Carbohydrate | g | 15.9 | |
| Fiber, total dietary | g | 6.5 | |
| Sugars, total | g | 9.36 | |
| Minerals | | | |
| Calcium, Ca | mg | 62 | |
| Iron, Fe | mg | 0.86 | |
| Magnesium, Mg | mg | 20 | |
| Phosphorus, P | mg | 19 | |
| Potassium, K | mg | 186 | |
| Sodium, Na | mg | 10 | |
| Zinc, Zn | mg | 0.17 | |
| Copper, Cu | mg | 0.095 | |
| Manganese, Mn | mg | 0.135 | |
| Vitamins | | | |

| Vitamin C mg 43.9 Thiamin mg 0.037 Riboflavin mg 0.09 Niacin mg 0.429 Pantothenic acid mg 0.208 Vitamin B-6 mg 0.036 Folate, total mcg 17 Folate, DFE 17 12 Vitamin A, IU IU 290 Vitamin A, RAE mcg_RAE 15 Vitamin E mcg_RAE 15 Taty acids, total saturated g 0.15 Fatty acids, total monounsaturated g 0.021 18:1 undifferentiated g 0.021 18:1 undifferentiated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Unity 155 Cryptoxanthin, beta mcg 155 Cryptoxanthin, beta mcg 155 | | | |
|---|------------------------------------|---------|-------|
| Riboflavin mg 0.09 Niacin mg 0.429 Pantothenic acid mg 0.208 Vitamin B-6 mg 0.036 Folate, total mcg 17 Folate, DFE mcg_DFE 17 Vitamin A, IU IU 290 Vitamin A, RAE mcg_RAE 15 Vitamin E mg 0.15 Lipids *** *** Fatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 155 | Vitamin C | mg | 43.9 |
| Niacin mg 0.429 Pantothenic acid mg 0.208 Vitamin B-6 mg 0.036 Folate, total mcg 17 Folate, DFE mcg_DFE 17 Vitamin A, IU IU 290 Vitamin E mcg_RAE 15 Vitamin E mg 0.15 Lipids Tatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.171 18:2 undifferentiated g 0.171 18:2 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Thiamin | mg | 0.037 |
| Pantothenic acid mg 0.208 Vitamin B-6 mg 0.036 Folate, total mcg 17 Folate, DFE mcg_DFE 17 Vitamin A, IU IU 290 Vitamin A, RAE mcg_RAE 15 Vitamin E mg 0.15 Lipids Fatty acids, total saturated g 0.154 felt undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Riboflavin | mg | 0.09 |
| Vitamin B-6 mg 0.036 Folate, total mcg 17 Folate, DFE mcg_DFE 17 Vitamin A, IU IU 290 Vitamin A, RAE mcg_RAE 15 Vitamin E mg 0.15 Lipids Tatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Niacin | mg | 0.429 |
| Folate, total meg 17 Folate, DFE meg_DFE 17 Vitamin A, IU IU 290 Vitamin A, RAE meg_RAE 15 Vitamin E mg 0.15 Lipids Fatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha meg 155 Cryptoxanthin, beta 190 | Pantothenic acid | mg | 0.208 |
| Folate, DFE mcg_DFE 17 Vitamin A, IU IU 290 Vitamin A, RAE mcg_RAE 15 Vitamin E mg 0.15 Lipids Fatty acids, total saturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta 15 | Vitamin B-6 | mg | 0.036 |
| Vitamin A, IUIU290Vitamin A, RAEmcg_RAE15Vitamin Emg0.15LipidsFatty acids, total saturatedg0.103Fatty acids, total monounsaturatedg0.15416:1 undifferentiatedg0.02118:1 undifferentiatedg0.137Fatty acids, total polyunsaturatedg0.17118:2 undifferentiatedg0.12418:3 undifferentiatedg0.047Othermcg155Carotene, alphamcg193 | Folate, total | mcg | 17 |
| Vitamin A, RAEmcg_RAE15Vitamin Emg0.15LipidsFatty acids, total saturatedg0.103Fatty acids, total monounsaturatedg0.15416:1 undifferentiatedg0.02118:1 undifferentiatedg0.137Fatty acids, total polyunsaturatedg0.17118:2 undifferentiatedg0.12418:3 undifferentiatedg0.047OtherCarotene, alphamcg155Cryptoxanthin, betamcg193 | Folate, DFE | mcg_DFE | 17 |
| Vitamin E mg 0.15 Lipids Fatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Vitamin A, IU | IU | 290 |
| Lipids Fatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Vitamin A, RAE | mcg_RAE | 15 |
| Fatty acids, total saturated g 0.103 Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Vitamin E | mg | 0.15 |
| Fatty acids, total monounsaturated g 0.154 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Lipids | | |
| 16:1 undifferentiated g 0.021 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Fatty acids, total saturated | g | 0.103 |
| 18:1 undifferentiated g 0.137 Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Fatty acids, total monounsaturated | g | 0.154 |
| Fatty acids, total polyunsaturated g 0.171 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | 16:1 undifferentiated | g | 0.021 |
| 18:2 undifferentiated g 0.124 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | 18:1 undifferentiated | g | 0.137 |
| 18:3 undifferentiated g 0.047 Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | Fatty acids, total polyunsaturated | g | 0.171 |
| Other Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | 18:2 undifferentiated | g | 0.124 |
| Carotene, alpha mcg 155 Cryptoxanthin, beta mcg 193 | 18:3 undifferentiated | g | 0.047 |
| Cryptoxanthin, beta mcg 193 | Other | | |
| | Carotene, alpha | mcg | 155 |
| Lutein + zeaxanthin mcg 129 | Cryptoxanthin, beta | mcg | 193 |
| | Lutein + zeaxanthin | mcg | 129 |

Table 2. Structural Features and Biological Activities of Polysaccharides

| Component name | Monosaccharide composition | Molecular weight (Da) | Glycosidic linkage | Chain conformation | Biological activities | References |
|----------------|---|-------------------------------------|---|--------------------------------------|---------------------------------------|---------------|
| FMPS | | 6.192 × 10 ⁶ (±2.59%) | β -glycosidic with a small amount of | Highly dispersive large polymer | Antioxidant activities. Antibacterial | [4, 8, 9, 23] |
| | | | α -glycosidic bonds | | activities | |
| FMPS1 | Galactose, glucose, galacturonic acid, rhamnose and mannose | 2.572×10^{7} (±0.517%) | α -glycosidic with a small amount of | Tight uniform spherical conformation | Pancreatic lipase active inhibition. | [4, 10] |
| | | | β -glycosidic bonds | | Bile acid-binding | |

| | | | | | abilities | |
|-------|---------------------------|-----------------------|----------------------|-----------------|---------------|---------|
| FMPS2 | Galactose, glucose, | 1.755×10^{6} | α -glycosidic | Random coil | Bile | [4, 10] |
| | galacturonic acid, | (±2.009%) | with a small | conformation | acid-binding | |
| | arabinose and mannose | | amount of | | abilities | |
| | | | β -glycosidic | | | |
| | | | bonds | | | |
| FMPS3 | Galactose, galacturonic | 2.563×10 ⁵ | β -glycosidic | Highly branched | Pancreatic | [4, 10] |
| | acid, arabinose, rhamnose | (±1.784%) | with a small | polymers | lipase active | |
| | and mannose | | amount of | | inhibition. | |
| | | | α -glycosidic | | Antioxidant | |
| | | | bonds | | activities | |
| FMPS4 | Galactose, galacturonic | 2.411×10^{5} | β -glycosidic | Highly branched | Antioxidant | [4, 10] |
| | acid, rhamnose and | (±1.808%) | linkage | polymers | activities | |
| | mannose | | | | | |

Table 3. Fragment Analysis of Limonin from F. margarita by Secondary Mass Spectrometry^[11]. Reprinted with Permission

| Order | Mass charge ratio (m/z) | MS/MS Fragment | Fragment ions affiliation | Structural formula |
|-------|-------------------------|--|---------------------------|--------------------|
| 1 | 515.2274 | C ₂₈ H ₃₅ O ₉ | $[M+H]^+$ | |

| 2 | 469.2205 | $C_{27}H_{33}O_{7}$ | $[M+H-CH_2O_2]^+$ | |
|---|----------|--|--|--|
| 3 | 455.2080 | $C_{26}H_{31}O_{7}$ | $[M+H-C_2H_4O_2]^+$ | |
| 4 | 437.1967 | $\mathrm{C}_{26}\mathrm{H}_{29}\mathrm{O}_{6}$ | $[\mathrm{M}+\mathrm{H}-\mathrm{C}_2\mathrm{H}_6\mathrm{O}_3]^+$ | |
| 5 | 411.2166 | $C_{25}H_{31}O_5$ | $[M+H-C_3H_4O_4]^+$ | |
| 6 | 205.0513 | C ₁₁ H ₉ O ₄ | $[M+H-C_{17}H_{26}O_5]^+$ | |

Table 4. Fragment Analysis of Nomilin from F. margarita by Secondary Mass Spectrometry^[11]. Reprinted with Permission

| Order | Mass charge ratio | MS/MS | Fragment ions affiliation | Structural formula |
|-------|-------------------|----------|---------------------------|--------------------|
| Order | (m/z) | Fragment | Fragment ions arimation | Structural formula |

| 1 | 471.2020 | $C_{26}H_{31}O_{8}$ | [M+H] ⁺ | |
|---|----------|---------------------|-------------------------------------|--|
| 2 | 453.1904 | $C_{26}H_{29}O_{7}$ | [M+H-H ₂ O] ⁺ | |
| 3 | 425.1966 | $C_{25}H_{29}O_6$ | $[M+H-C_2H_4O_2]^+$ | |

Structural Features and Biological Activities of Bioactive

Compounds from Fortunella margarita (Lour.) Swingle: A Review

ZENG Hong-Liang(曾红亮) CHEN Pei-Lin(陈培琳) HUANG Can-Can(黄灿灿) SHEN Jin-Ye(沈瑾烨)

CHANG Qing(常 青) ZHENG Bao-Dong(郑宝东) ZHANG Yi(张 怡)

This review focuses on the structural features and biological activities of polysaccharides, limonoids, essential oils and flavonoids and other bioactive substances from *F. margarita* and their potential applications in food, daily chemical and pharmaceutical industries. The polysaccharides were a macromolecular heteropolysaccharide, containing four kinds of polysaccharide fractions with different molecular weights. Different polysaccharide fractions displayed antibacterial, antioxidant activities and hypolipidemic effect. This functionality was affected by the monosaccharide composition, glycosidic linkage, molecular weight and chain conformation in aqueous.

